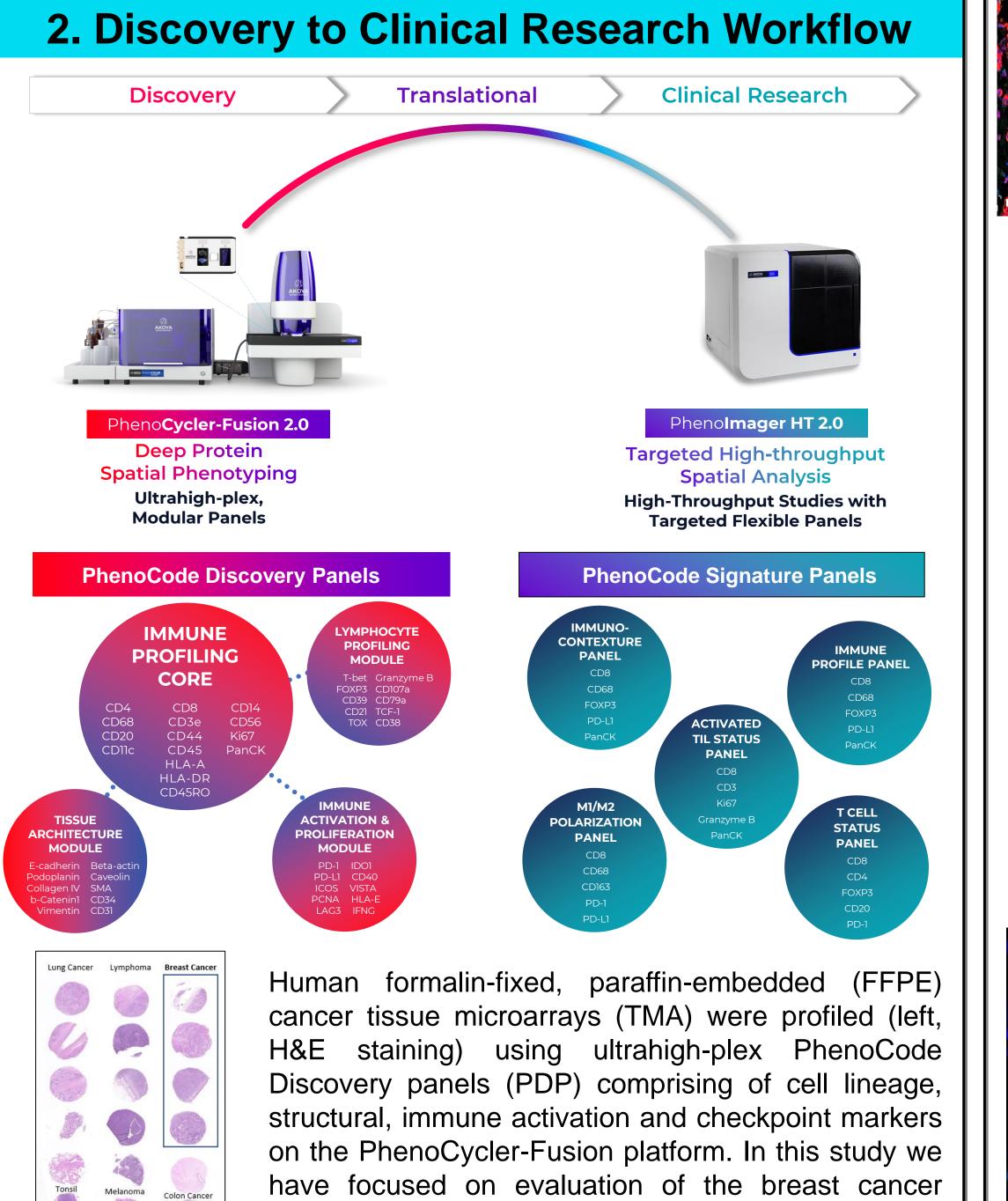


1. Introduction

The transition from ultrahigh-plex discovery experiments in multiplexed imaging to targeted, high-throughput translation and thereby clinical research studies presents a significant challenge. Successful translation hinges on the harmonization of staining, imaging, and data analysis technologies across platforms. This work aims to present a streamlined workflow for the transition, leveraging the capabilities of the PhenoCycler®-Fusion 2.0 and PhenoImager® HT platforms.

We demonstrate a two-step approach utilizing PhenoCode[™] panels: 1. Deep Protein Spatial Phenotyping: For this step, tumor microarrays (TMAs) comprised of human formalin-fixed, paraffin-embedded (FFPE) cancer tissues were phenotyped using the ultra-high-plex PhenoCode Discovery panels on the PhenoCycler-Fusion 2.0 platform. This step allows for extensive characterization of the tissue microenvironment at the protein level. 2. Targeted High-Throughput Analysis: Following discovery, PhenoCode Signature panels were employed on the PhenoImager HT 2.0 platform. This platform enables highthroughput analysis of protein targets identified during the discovery phase, facilitating the translation of these findings into targeted translational and clinical research studies.



subtypes in the TMA by running sequential PhenoCode Signature panels (PSP) using the PhenoImager HT platform targeted to key biomarkers of immune contexture, T cell activation status and macrophage polarization identified from the discovery studies.

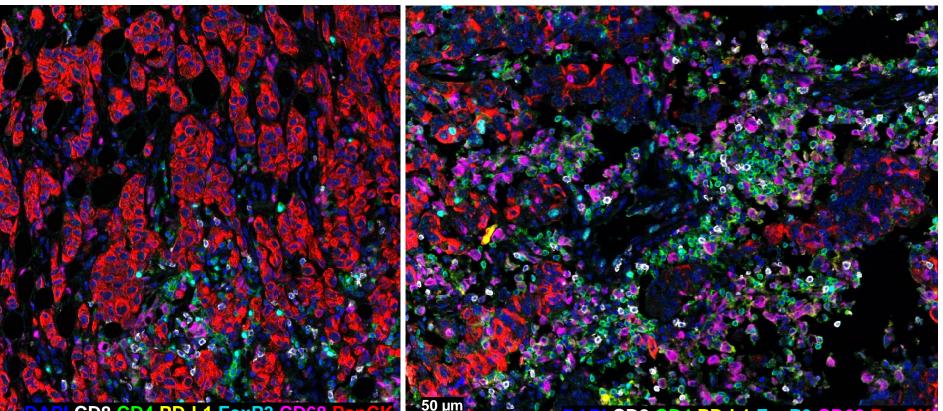
5504: Integrating ultrahigh-plex spatial phenotyping: From discovery to clinical applications

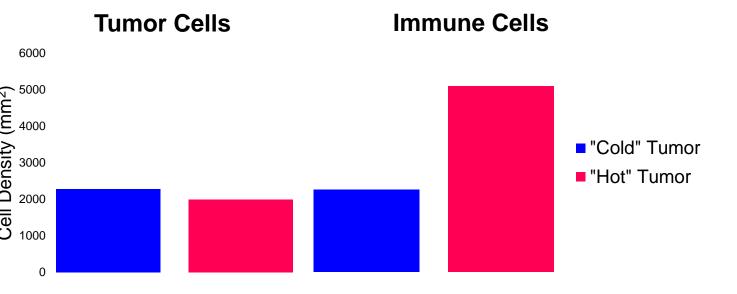
3. Ultrahigh-Plex Spatial Phenotyping of Breast Cancer Tissues

Single-Cell Spatial Phenotyping of Proteins in Breast Cancer Tissues using 32-plex Discovery Panel reveals distinct cell populations. Three types of breast cancer TMA cores: triple negative breast cancer, invasive breast carcinoma, and invasive ductal carcinoma were imaged and analyzed. Heatmap (A) corresponding to average expression of cell-type defining markers revealed 13 distinct cellular populations identified by unsupervised clustering. (B) Representative images from invasive ductal carcinoma. The difference in the immune cell composition and status in the invasive ductal carcinomas were further analyzed using 6-plex PhenoCode Signature Panels which are ideally suited for automated high-throughput studies

4. Quantitative Spatial Profiling of Breast Cancer Subtypes using 6-plex Multiplex Imaging and Phenotyping Analysis

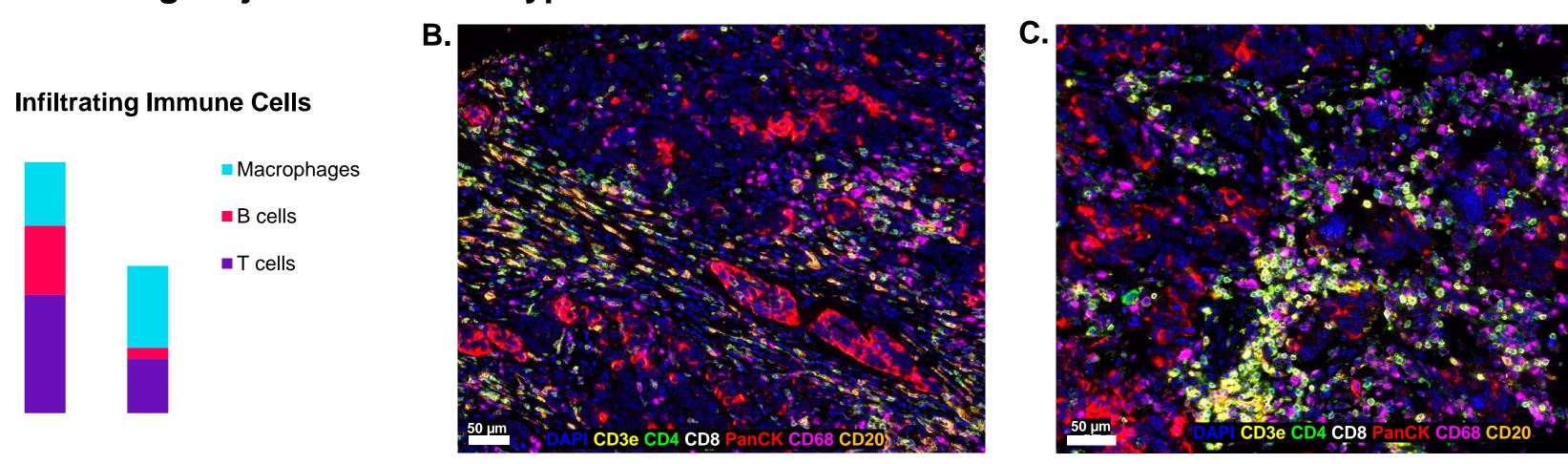
4.1. Assessing immune cells infiltration in the tumor





4.1. Is the tumor "hot" or "cold"? The PSP Immuno-Contexture panel + CD4 can be used to identify the specific immune cell types to quickly determine if a tumor is "hot" or "cold". Despite comparable numbers of tumor cells, the immune cell populations are enriched within the invasive ductal carcinoma sample relative o the triple negative breast cancer sample, indicating a "hot" tumor

4.2. Determining major immune cell types in the tumor

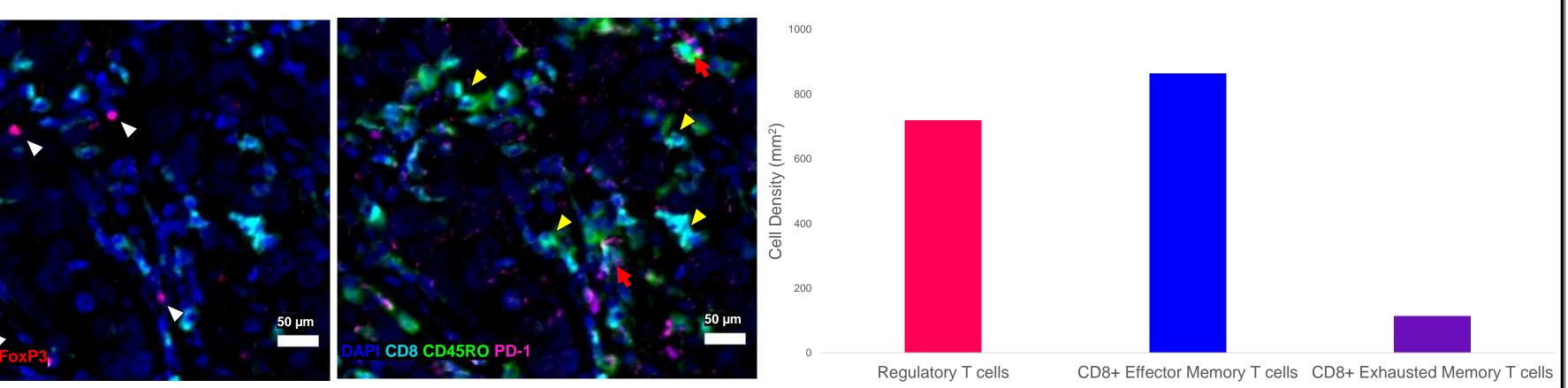


4.2. Assessment of major immune cell types in the tumor. The PSP Immune Profile panel + CD4 was used to quantify the major immune cell types in the different invasive ductal carcinoma samples. Assessment with the Immune Profile panel + CD4 showed differences in macrophage, B and T cell densities between the invasive ductal carcinoma samples (A). Representative images of the Immune Profile panel from the different samples are shown (B

4.3. Analysis of T cell status within the tumor

80%

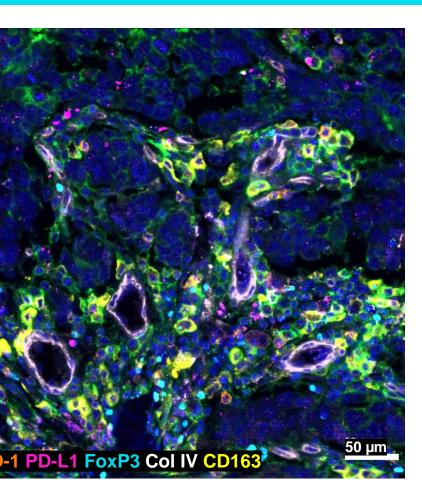
60%



4.3. Determination of T cell status. The PSP T Cell Status panel + CD45RO was used to phenotype exhaustive T cells. Regulatory T cells (CD8-/FoxP3+ white arrowhead – left image), CD8 Effector Memory T cells (CD8+/CD45RO+/PD-1-, yellow arrowhead – center image), and CD8 Exhausted Memory T cells (CD8+/CD45RO+/PD-1+, red arrow – center image) indicate the status of the T cells in an invasive ductal carcinoma sample. Cell density for the sample is shown on the right.

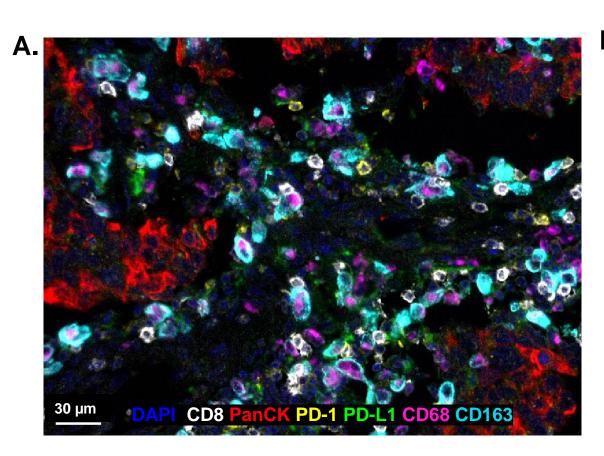
Ning Ma¹, Katie Miller¹, Aditya Pratapa¹, Niyati Jhaveri¹, and Agnes Haggerty¹ ¹Akoya Biosciences, Marlborough, Massachusetts, USA

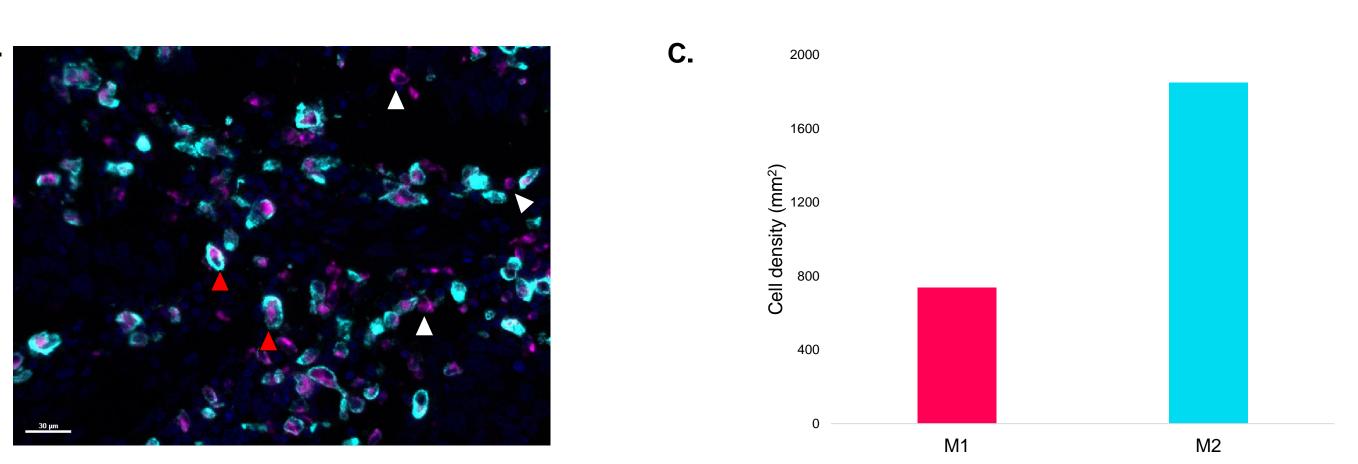






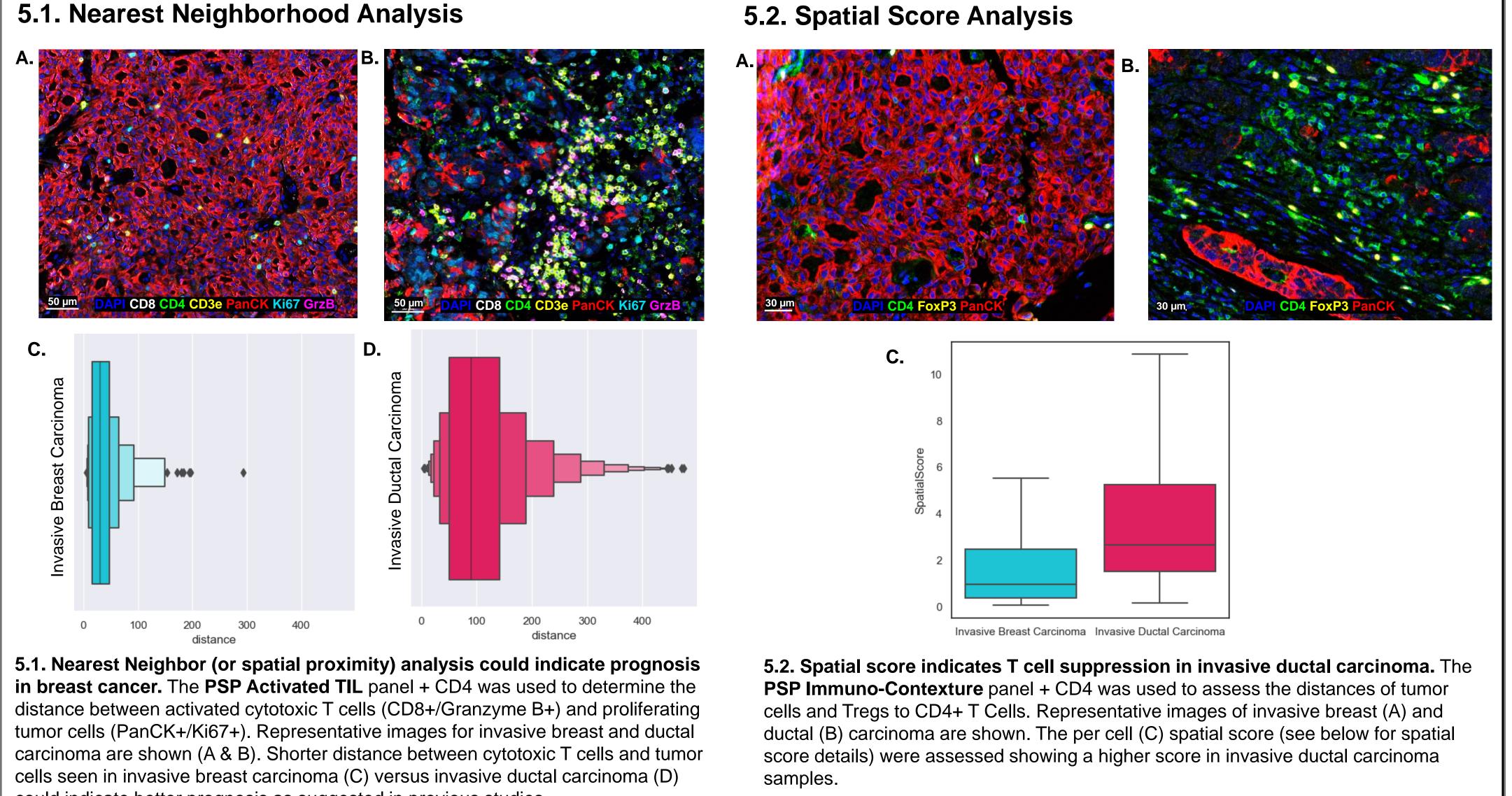
4.4. Assessing macrophage polarization in the tumor



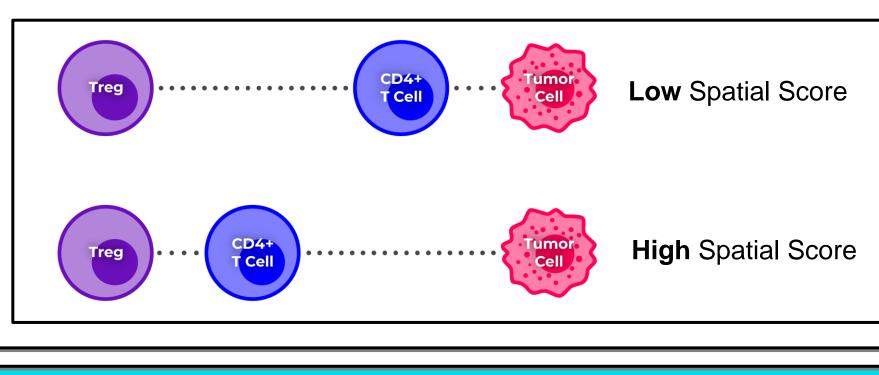


4.4. Single-cell spatial phenotyping of invasive ductal carcinoma reveals distinct macrophage polarization profiles. The PSP M1/M2 Polarization panel + PanCK was used following the Immune Profile panel to assess macrophage polarization in an invasive ductal carcinoma core. Representative images are shown. (A) 6-plex and (B) M1 (CD68+/CD163-, white arrowhead) & M2 (CD68+/CD163+, red arrowhead). Cell densities for the M1 and M2 macrophages are shown on the right (C).

5. Assessment of Spatial Relationships in Breast Cancer Subtypes



could indicate better prognosis as suggested in previous studies.



6. Unlocking the Power of PhenoCode Panels for Ultrahigh-plex Discovery to Targeted High-throughput Studies

PhenoCode panels, powered by Akoya's novel barcode chemistry, provide an off-the-shelf solution immunofluorescence staining that requires minimal optimization. Our study showcases how ultrahigh-plex profiling using PhenoCode Discovery panels can be followed by a more targeted analysis using PhenoCode Signature panels. Here, we have shown how the PhenoCode Signature panels can be leveraged for rapid immune profiling and immune status assessment of the TME and to perform spatial analysis to gain key biological insights that can be used for the development of spatial signature that can more reliably predict immunotherapy response. akoyabio.com/phenocode/

The Spatial Score calculated here is the ratio of the physical distance between CD4+ T-cells and the nearest tumor cell, relative to its nearest Treg (CD4+ T Cell to Tumor Cells / CD4+ T Cell to Tregs). Tregs can exert immunosuppressive effects on CD4+ T-cells, which then results in a higher Spatial Score.

Combining these spatial analyses can provide a more in depth understanding of the TME. The invasive breast carcinoma displays a shorter distance by spatial proximity measurement than the invasive ductal carcinoma (5.1), which correlates with the spatial score results (5.2)

