

Cell Profiling Core	CD4, CD68, CD20, CD11c, CD8, HLA-DR, Ki67, CD45RO, CD3ε, PanCK, CD44, CD45, HLA-A, CD14, CD56 + CD163, CD11b, CD57
Lymphocyte Profiling Module	CD21, FOXP3, Granzyme B, CD79α, TCF-1, CD38, TOX, T-bet, CD107α, CD39
Immune Activation and Proliferation Module	PD-1, PD-L1, IDO1, LAG3, VISTA, IFNG, HLA-E, CD40, ICOS, PCNA
Tissue Architecture Module	E-cadherin, CD31, Podoplanin, SMA, Vimentin, Collagen IV, CD34, β -catenin1, β -actin, Caveolin
Custom Metabolic Module	ASCT2, CPTIA, HK1, LDHA, IDH2, GLUTI, pNRF2, ATP5A, Citrate Synthase, HIF1α, G6PD, SDHA

The Spatial Biology Company[™]

6872: A Spatio-Temporal Approach to Mapping the Dynamics of Cutaneous Squamous **Cell Carcinoma Progression and Immunotherapy Response: A Journey through TiME**

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- Cellula Phenotypes (colored as shown the right) on hierarchica expression of cell lineage and structural markers from the PhenoCode Discovery Panels
- t-SNE Plot showing the data and distinct cellular phenotypes in reduced dimensionality.

A) Whole-Slide Multiplex PCF Images of a cSCC, labeled with (E-cadherin), vascular CollV), immune (CD45), proliferation (Ki67) markers at baseline and 6 months post neterogeneity landscapes across the course o treatment in the same patient.

Single-Cell Spatial Maps illustrate the differences in the spatia the aanization of twelve listinct cellular phenotypes (legend in section #3.1). A marked difference is observed in overal tumor burden.

Charts illustrate the of the different phenotypes. Higher percentages of M1 macrophages (orange), CD8+ T cells (dark purple) and plasma cells (brown) are seen post treatment, correlating with regression of the tumor (pink).

A) Whole-Slide Multiplex PCF Images of a cSCC, labeled with epithelial (E-cadherin), vascular (CD31, CollV), immune (CD45), (GLUTI) metabolic and proliferation (Ki67) markers at baseline and 12 months post reveal the heterogeneity spatia landscapes across the course of

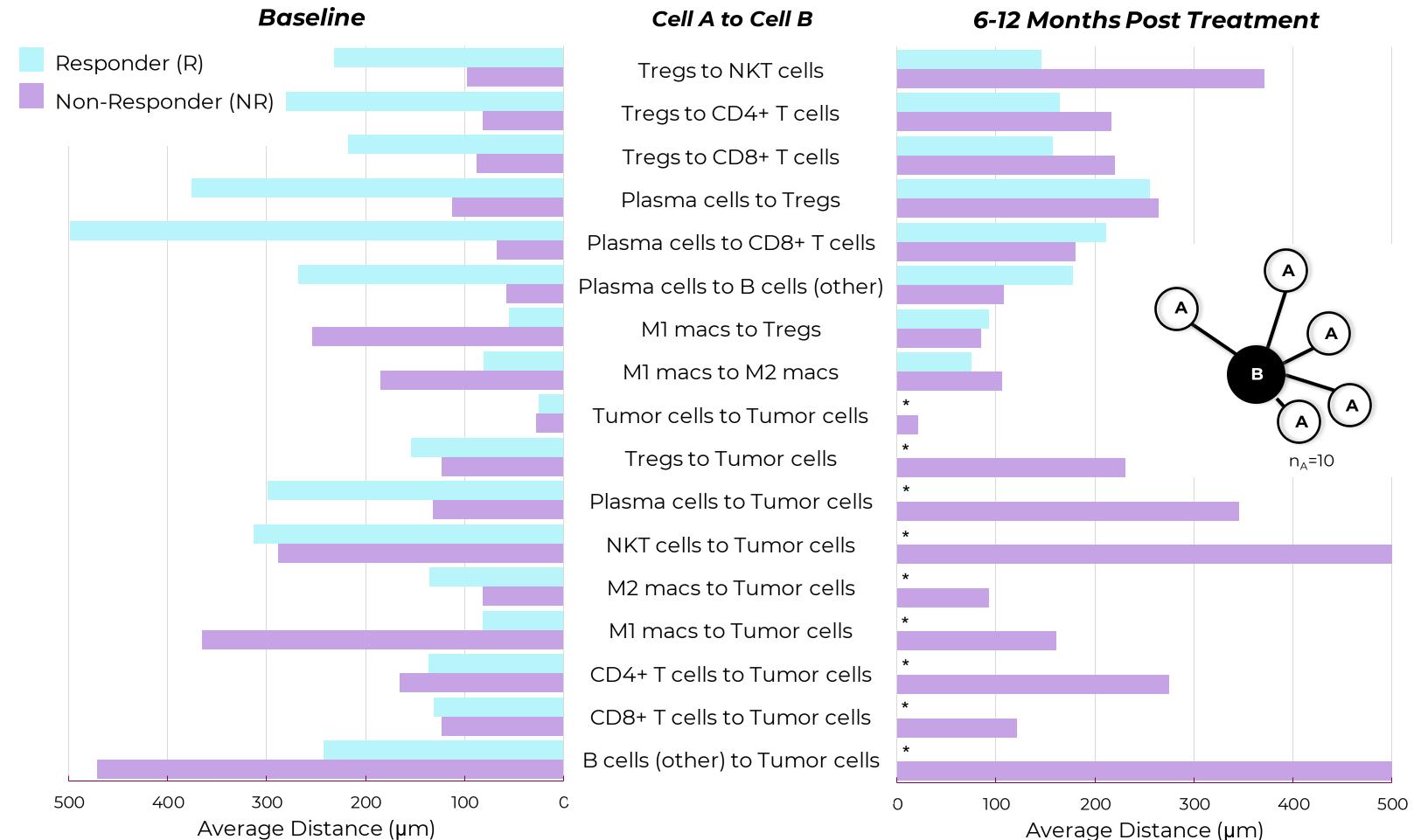
treatment in the same patient

Single-Cell Spatial Maps illustrate the differences in the spatial organization of the twelve distinct cellular phenotypes (legend in section #3.1). A marked increase is observed in overall tumor burden.

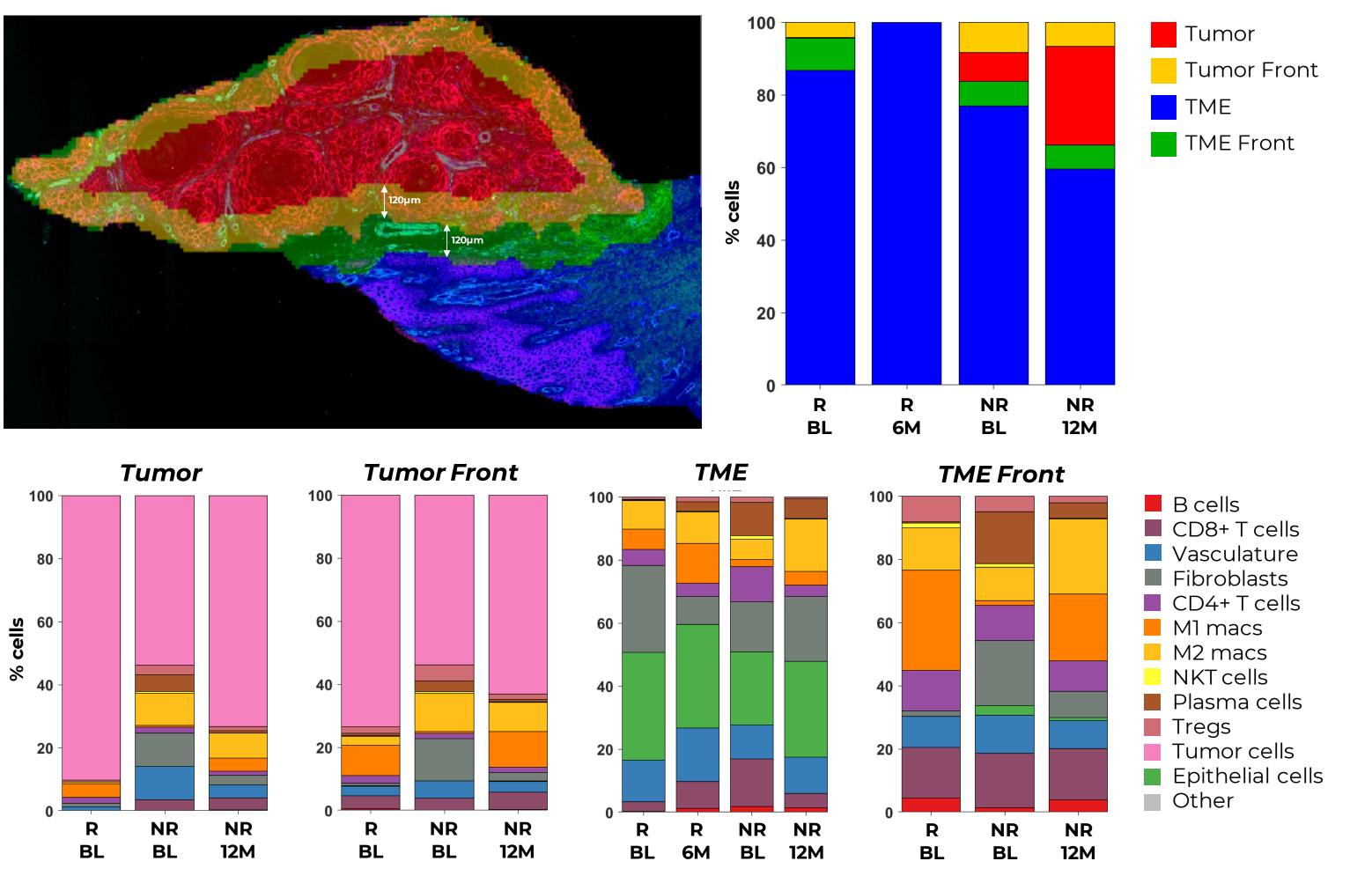
C) Bar Charts illustrate the abundance of the different phenotypes. Higher percentages of M2 macrophages (mustard yellow) along with a decrease i CD8+ T cells (dark purple) and plasma cells (brown) post treatment correlates with tumor proliferation (pink).

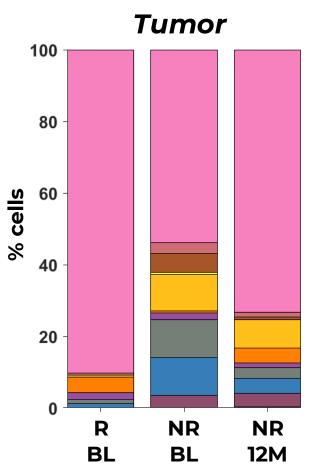






Spatial Proximity Analysis shows the localization of cellular phenotypes relative to each other by measuring distances of 10 neighboring "A" cells from a focal "B" cell. Differences in cellular distribution emerge between the responder and non-responder at baseline as well as temporally within the same patient (* denotes not applicable).





Tissue Compartmental Analysis highlights regional heterogeneity in cellular composition within the tumor, at the tumor front, in the tumor microenvironment (TME) and at the TME front. Most notably, differences are observed in M1 macrophages, M2 macrophages and plasma cells at baseline (BL) and 6-12 months (6-12M) post treatment across responder (R) and non-responder (NR) cSCC samples.

Cancer development is a dynamic process, characterized by cellular and molecular changes in the tissue microenvironment that contribute to sustained proliferation, immune evasion, and resistance. Since cSCC biopsies can be collected non-invasively, longitudinal studies tracking spatio-temporal changes can follow the course of the tumor in the same patient. The PCF 2.0 platform is ideally suited for such studies with its high plex, high throughput single-cell resolution, seamless scaling, and unparalleled high plex, high throughput single-cell resolution, seamless scaling, and unparalleled speed

4. Spatial Analyses Reveals Regional Heterogeneity

4.1 Spatial Proximity Analyses Reveals Temporal Differences in Cellular Distribution



5. The Power of Spatio-temporal Mapping at Scale

