

## 1 Background

Understanding cellular heterogeneity and spatial relationships between biomarkers within the tumor microenvironment is a key component to translational research in immuno-oncology. A reproducible, quantitative, easy-to-use, and standardized multiplex fluorescent IHC assay is required for quantitative assessment of these relationships *in situ* for current I/O clinical trials and translational researches. In this study, we demonstrate a fully developed, yet flexible, end-to-end workflow solution for tissue biomarker discovery in lung cancer and melanoma. This newly developed Phenoptics™ solution provides an integrated MOTiF™ workflow including optimized RTU reagents plus image analysis algorithms enabling a more comprehensive and specific tumor microenvironment analysis with minimal user tuning.

## 2 Methods

FFPE samples from human lung cancer and melanoma were stained using MOTiF PD1/PD-L1 Panel: Auto LuCa Kit and MOTiF PD1/PD-L1 Panel: Auto Melanoma Kit. Staining was performed on the Leica BOND RX™ automated stainer with the preloaded MOTiF protocol. Multispectral scans were acquired on Vectra Polaris® with pre-optimized acquisition parameters and analyzed with a pre-configured phenotyping algorithm in inForm®. Spatial analyses and visualizations were performed in R using phenoptr and phenoptr Reports [1].

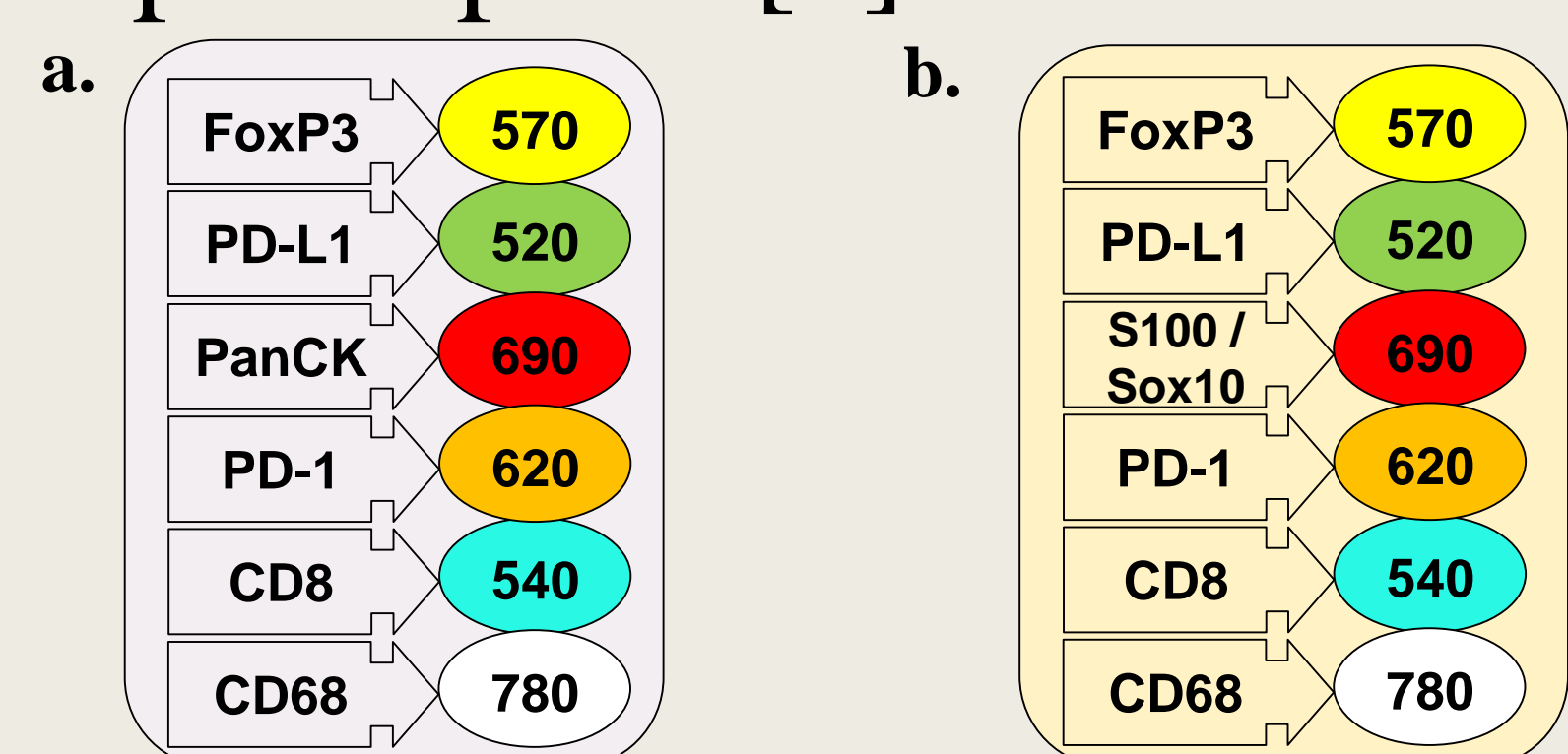


Fig. 1 Antibody-Opal pairs for MOTiF PD1/PD-L1 Panel: Auto LuCa Kit (a) and MOTiF PD1/PD-L1 Panel: Auto Melanoma Kit (b).

## 3 MOTiF Panel Kit Workflow



**MOTiF Panel Kit** Quality Reagents | **BOND RX** Optimized protocol | **Vectra Polaris** Standardized imaging protocol | **InForm Software** Pre-configured analysis algorithms

Fig. 2 Akoya new 7-color MOTiF PD1/PD-L1 Panel Kits along with whole slide multispectral imaging workflow provide the only validated end-to-end solution for unparalleled quantitative data for translational immuno-oncology research.

## 4 Results

### Pathologist-verified Antibody Performance

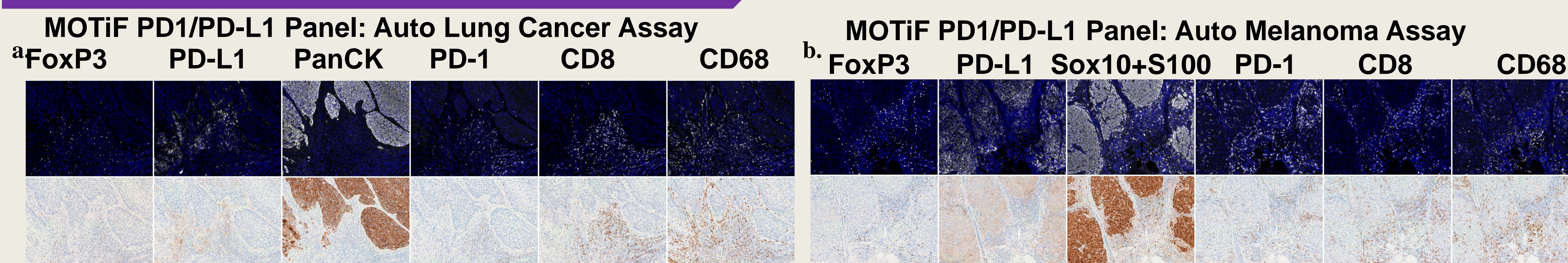


Fig. 4 Chromogenic and Fluorescence Concordance Matrix for MOTiF PD1/PD-L1 Panel Assay. Consecutive sections from lung cancer and melanoma are stained with the MOTiF PD1/PD-L1 Panel Kits or Leica BOND Polymer Refine Detection Kit. Representative Fields comparing DAB and Unmixed Monoplex view from the MOTiF PD1/PD-L1 Panel Assay from Lung Cancer (a) and Melanoma (b).

### Rules-based Phenotyping Analysis

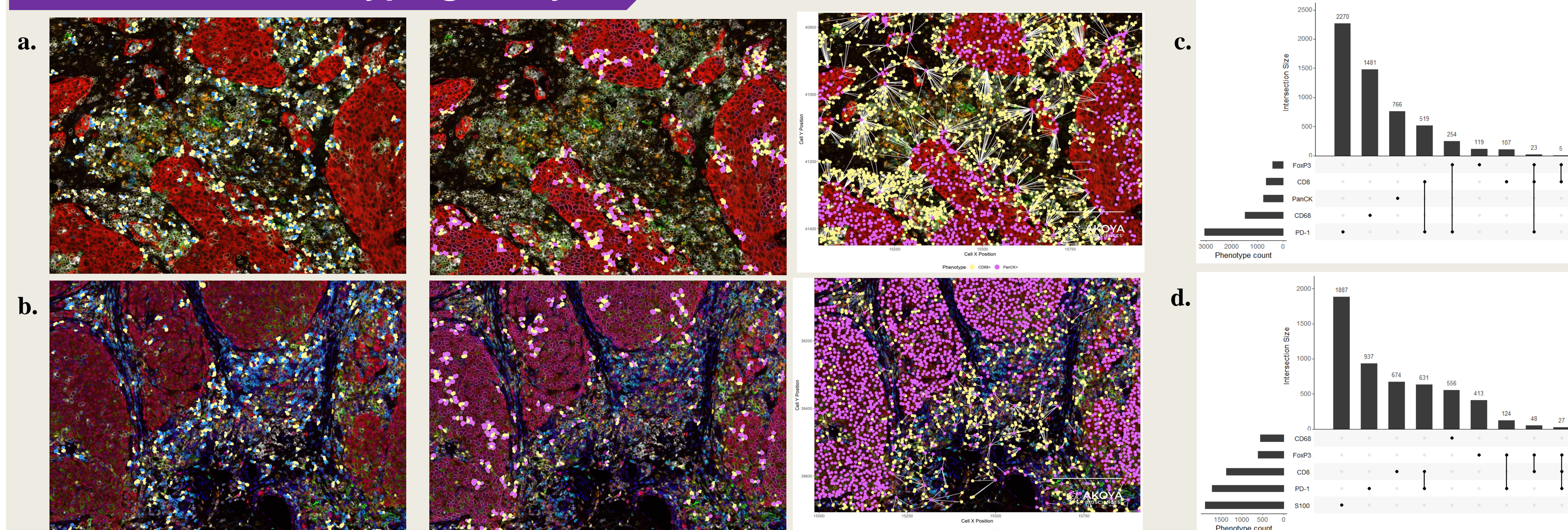


Fig. 5. Touching cell (left and middle) and nearest neighbor (right) plots from phenoptrReports for lung cancer (a) and melanoma (b). Each plot shows two phenotypes (CD8+ is blue, CD68+ is yellow, and tumor+ is pink). In touching cell plots, a cell outline is filled if it is touching a cell of the paired phenotype. In nearest neighbor plots, the nearest PanCK+ cell to each CD8+ cell is connected by a white line. (c) and (d) are summaries of cell phenotype counts for the fields in (a) and (b), respectively. Horizontal bars show counts of individual positivities. The vertical bars show counts of specific phenotype combinations present in the data. The central matrix shows the combinations graphically.

### Reproducibility of MOTiF PD1/PD-L1 Panel Kits

Tissue	CD8 <sup>+</sup> PD-L1 <sup>+</sup> PD-1 <sup>+</sup> FoxP3 <sup>+</sup> PanCK <sup>+</sup> CD68 <sup>+</sup>					
	Opal Polaris 480	Opal 520	Opal 570	Opal 620	Opal 690	Opal Polaris 780
Lung Cancer	15.0	12.2	15.9	12.7	9.5	13.3
Melanoma	16.7	9.8	14.5	14.8	14.4	16.3

Fig. 6 Top Quartile results from Phenotyping analysis on serial section slides from 3 different tissues stained with MOTiF PD1/PD-L1 Panel kits. Phenotyping algorithm applied to all tissues and top 25% of positive cells reported. CV's calculated by taking the mean and standard deviation of individual fields across five different slides (StDev/Mean \* 100). CV's were then averaged between fields (lower table). A scoring threshold was used to determine PD-L1+ cells for analysis.

## 7-Color Whole Slide Multispectral Imaging

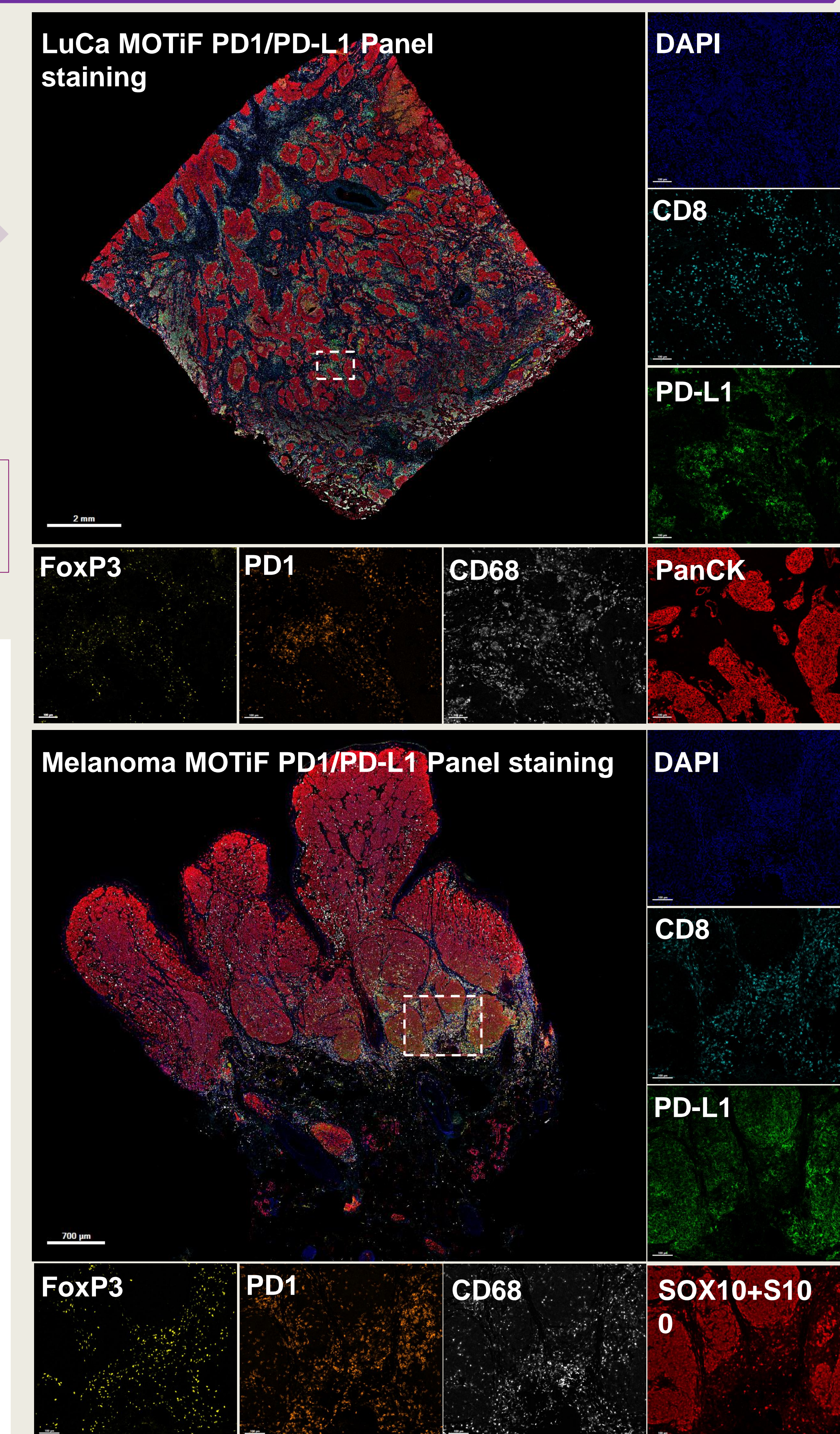


Fig. 3 Whole slide scan of MOTiF PD1/PD-L1 Panel assay on lung cancer (upper) and melanoma (lower) samples.

## 5 Conclusions

With new MOTiF PD1/PD-L1 Panel Kits, we have demonstrated an easy-to-use yet comprehensive end-to-end Phenoptics research workflow. We have radically simplified the Opal method and facilitated the development and optimization of translational multiplex fluorescent assays by providing pre-defined staining conditions while still giving researchers the flexibility to balance signals based on their tissue samples. Complementary pre-configured imaging protocol and analysis algorithm provide researchers faster access to quantitative data across study samples.