

6-Plex, 7-Color MOTiF™ Mouse Tissue Analysis Protocol

GETTING STARTED

The following protocol has been optimized for use with the Leica BOND RX and the Opal Polaris 7 Color Automation IHC Detection Kit. Stained slides can be imaged using

the Vectra Polaris imaging system. Currently, this kit is not compatible with Vectra 3. Additional components are required for imaging on the Mantra Workstation.

MATERIALS

	Slides	Catalog #	Contents
Opal Polaris 7 Color Automation IHC Detection Kit	50 Slides	NEL871001KT	<ul style="list-style-type: none"> • 1X Plus Automation Amplification Diluent (2 x 50mL) • Opal Polaris 480 Reagent • Opal 520 Reagent • Opal 570 Reagent • Opal 620 Reagent • Opal 690 Reagent • Opal Polaris 780 Reagent <ul style="list-style-type: none"> - Opal TSA-DIG - Opal Polaris 780 • Spectral DAPI solution (1.5mL) • DMSO (500uL) • Blocking/ Ab Diluent (2 x 100mL) • Opal Polymer HRP Ms+Rb (2 x 50mL)
Opal Polymer anti-Rabbit HRP Kit	50 Slides	ARR1001KT	<ul style="list-style-type: none"> • Opal Polymer Anti-Rabbit HRP (10mL) <ul style="list-style-type: none"> - Recommended working dilution (1:5) • Opal Polymer Anti-Rabbit HRP Diluent (40mL)

*NOTE: All dispensing volumes will be at 150µL per slide, with double dispenses for Opals and DAPI.

ADDITIONAL REQUIRED MATERIALS AND REAGENTS

These materials are not included in the kit and must be supplied separately

Laboratory Materials	BOND RX Materials
<ul style="list-style-type: none"> • Histological grade ethanol (for rehydration) • Tris-buffered saline (TBS) wash buffer <ul style="list-style-type: none"> - (25 mM TRIS-HCl; pH 7.5 150 mM NaCl) • Peroxidase-free water <ul style="list-style-type: none"> - This specification may be met by commercial "cell culture grade" water or ultra-pure (i.e. Milli-Q™) water that has been autoclaved • Mounting medium • Glass coverslips 	<ul style="list-style-type: none"> • Titration Kit (OPT9049) • Open container - 7mL (OP79193) • Open container - 30mL (OP309700) • Research Detection System 2 (DS9777) • Universal Covertiles (S21.4611) • Slide Tray (S21.0304) • Reagent Tray (S21.1003) • Slide Labels and Printer Ribbon (S21.4564) • Apex Adhesive Slide (3800040) • Dewax Solution (AR9222) • Epitope Retrieval Solution 1 (AR9961) • Epitope Retrieval Solution 2 (AR9640) • Wash Solution 10X Concentrate (AR9590) • Aspirating Probe Cleaning System (CS9100)

SOLUTION PREPARATION

Primary Antibody Working Solution

Dilute primary antibody in the Antibody Diluent/Block at optimal concentration for Opal detection as recommended in the chart on page 4.

Secondary Antibody Working Solution

Using the contents of the Opal Polymer anti-Rabbit HRP Kit, create a 1:5 Working Solution of the Opal Polymer Anti-Rabbit HRP (10mL) by combining it with the Opal Polymer Anti-Rabbit HRP Diluent (40mL). For this protocol, the Opal Polymer HRP Ms+Rb supplied in the Opal Polaris 7-color Automation IHC Detection Kit will not be used.

Opal Working Solution*

Reconstitute each Opal reagent in 75µL of DMSO, with the exception of Opal Polaris 780 (see below). Before each procedure, dilute Opal reagent in 1X Plus Automation Amplification Diluent to make Opal reagent working solution. We recommend to start diluting the Opal reagent at 1:150. Optimize your assay according to the Opal Assay Development Guide.

Opal Polaris 780 Working Solution*

Reconstitute TSA-DIG in 75µL of DMSO, and Opal Polaris 780 in 300µL of deionized water. Before the procedure, dilute TSA-DIG in 1X Plus Automation Amplification Diluent at 1:100 to make TSA-DIG working solution. Dilute Opal Polaris 780 with Antibody Diluent/ Blocking at 1:25 to make the working solution.

DAPI Working Solution*

Add 2-3 drops of DAPI solution into 1mL of TBS or TBST. Approximately 300µL of DAPI Working Solution is required per slide. Discard any unused portion of DAPI Working Solution.

***NOTE:** To help assist in your Working Solutions calculations, one dispense equals 150µL of Working Solution. For this assay, two dispenses of each Opal reagent and TSA-DIG are recommended per slide (with the exception of Opal Polaris 780) to achieve optimal staining, plus additional solution for the dead volume of the container. Discard any unused portion of any of the Working Solutions when the run is complete.

BOND RX Wash Solution

Create a working 1X BOND RX Wash Solution by diluting the stock 10X concentrate BOND™ Wash Solution with peroxidase-free water.

SPECIAL CONSIDERATIONS AND BOND RX PROTOCOL

Opal Polaris 780 Automation Steps

The Opal Polaris 780 reaction is antibody based. Because of this, there must be additional washing steps to cool down the slide between the TSA-DIG stripping step (with ER1 at 95°C) and before the Opal Polaris 780 application step.

NOTE: The Opal Polaris 780 must ALWAYS go last in your multiplex.

BOND RX Wash Solution Steps

In the following protocol, users will see the following step: BOND Wash Solution. This refers to a series of wash steps that are built-in to the Leica protocol. Listed below are a breakdown of those steps:

- **Post-Primary Antibody, Epitope Retrieval, Opal 780, and DAPI application:**
0 min, 1 min, 0 min
- **Post-Opal Polymer HRP application:**
0 min, 1 min, 3x 0 min
- **Post-Opal application and TSA-DIG (except Opal 780):**
0 min, 1 min, 2x 0 min

DIRECTIONS

Follow the steps listed below to perform the automated multiplex immunofluorescence staining assay.

Differs from standard Opal BOND RX protocol.

STEP 0 BOND RX Protocol Creation

- a. Create a copy of the *Opal 7-color (v5.2 plus) IHC protocol.
- b. Alter the copied version to reflect the protocol beginning at Step 2.

STEP 1 Slide Preparation

- a. Bake the FFPE slides for 3 hours at 65°C in a laboratory oven.
- b. When setting up the BOND RX Study Protocol, select the following listed settings for Preparation and HIER (see Figure 1).
- c. Label the slides with their corresponding BOND RX barcode.
- d. Load the slides into the BOND RX trays, and prepare and load the reagents.
- e. Press “Start” to begin the BOND RX protocol..

STEP 2 Research Detection System

- a. TBS wash, 0 min

STEP 3 Blocking

- a. Akoya blocking buffer, 5 min

STEP 4 Primary Antibody Incubation

- a. Antibody 1 incubation, 30 min
 - i. Use the chart below to determine antibody information, staining order, and dilution factor.

The screenshot shows the BOND RX software interface with the following settings:

- Staining mode:** Single (dropdown), Routine (dropdown)
- Process:** IHC (selected), ISH
- Marker:** *Negative (dropdown)
- Staining:** Opal 7 Color MOTIF (dropdown)
- Preparation:** *Dewax (dropdown)
- HIER:** *HIER 40 min with ER2 (dropdown)
- Enzyme:** *--- (dropdown)

Order	Antibody	Clone	Vendor	Species	AB Dilution Factor	Opals	Opal Dilution Factor
1	CD8a	D4W2Z	Cell Signaling	rabbit mAB	1:200	Opal 480	1:150
2	Bcl-2	EPRI7509	Abcam	rabbit mAB	1:500	Opal 520	1:150
3	CD4	D7D2Z	Cell Signaling	rabbit mAB	1:50	Opal 570	1:100
4	F4/80	D2S9R	Cell Signaling	rabbit mAB	1:100	Opal 620	1:400
5	CD31	D8V9E	Cell Signaling	rabbit mAB	1:300	Opal 690	1:150
6	Ki-67	IHC-00375	Bethyl	rabbit pAB	1:150	TSA-DIG, Opal 780	1:100, 1:25

STEP 5 Opal Polymer HRP

- a. Bond Wash Solution
- b. Opal Polymer Anti-Rabbit HRP, 10 min

STEP 6 Signal Amplification

- a. Bond Wash Solution
- b. Double dispense the Opal fluorophore (Use the chart to determine Opal pairing and dilution factor)
 - i. First dispense, 0 min
 - ii. Second dispense, 10 min

STEP 7 Antibody Stripping

- a. Bond Wash Solution
- b. Bond ER solution 1
 - i. Temperature at 95°C, incubate at 20 min
- c. Bond Wash Solution

STEP 8 Repeat Steps 3-7

- a. Repeat Steps 3-7 for the next four antibodies.
- b. Go on to Step 9 for Opal Polaris 780 staining.

STEP 9 Blocking

- a. Akoya blocking buffer, 5 min

STEP 10 Primary Antibody Incubation

- a. Antibody 6 incubation, 30 min

STEP 11 Opal Polymer HRP

- a. Bond Wash Solution
- b. Opal Polymer Anti-Rabbit HRP, 10 min

STEP 12 Introduction of Opal TSA-DIG

- a. Bond Wash Solution
- b. Opal TSA-DIG [1:100]#
 - i. First dispense, 0 min
 - ii. Second dispense, 10 min

STEP 13 Antibody Stripping

- a. Bond Wash Solution
- b. Bond ER Solution 1
 - i. Temperature at 95°C, incubate at 20 min
- c. Bond Wash Solution#
 - i. Wash
 - ii. 2X Wash, 10 min incubations each
 - iii. Wash

STEP 14 Opal Polaris 780 Signal Generation

- a. Opal Polaris 780 [1:25], 60 min#
- b. Bond Wash Solution

STEP 15 DAPI Counterstain and Mount

- a. Spectral DAPI#
 - i. Open dispense, 0 min
 - ii. Open dispense, 5 min
- b. Bond Wash Solution

MULTIPLEX ASSAY PANEL TABLE

Use the table provided below to keep track of your assay panel development.

Project Name: _____

Date: _____ Tissue(s): _____

Researcher: _____

Order	Antibody	Supplier	Clone/ Lot	Category #	Dilution Factor	Opal Pairing	AR
1							
2							
3							
4							
5							
6							
7							
8							

Notes: _____

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